

Supplemental material: Stable isotope analysis of vertebrae reveals ontogenetic changes in habitat in an endothermic pelagic shark

Aaron B. Carlisle^{1*}, Kenneth J. Goldman², Steven Y. Litvin¹, Daniel J. Madigan¹, Jennifer S. Bigman³, Alan M. Swithenbank¹, Thomas C. Kline Jr.⁴, Barbara A. Block¹

¹ Hopkins Marine Station of Stanford University, 120 Oceanview Blvd, Pacific Grove, CA, United States

² Alaska Department of Fish and Game, 3298 Douglas Place, Homer, AK, United States

³ Moss Landing Marine Laboratories, 8272 Moss Landing Rd., Moss Landing, CA, United States

⁴ P.O. Box 1949, Cordova, AK, United States

*Corresponding author: aaroncar@stanford.edu

Methods

Between 200–600 µg of decalcified tissue was weighed into tin boats (4 x 6 mm, Costech) and analyzed at the Stable Isotope Laboratory at University of California Santa Cruz using an Elemental Analyzer coupled to an isotope ratio mass spectrometer (Delta XP-EA, Thermo-Finnagen IRMS). Isotopic composition is expressed using δ notation, which is calculated using the equation: $\delta X = [(R_{\text{sample}}/R_{\text{standard}})-1] * 1000$; where X= ¹³C or ¹⁵N, R= ratio of ¹³C/¹²C, ¹⁵N/¹⁴N, and the standards are Vienna Pee Dee Belemnite limestone (V-PDB) for carbon and AIR for nitrogen. Analytical precision, based on an internal lab standard of known carbon and nitrogen isotopic composition (n = 106), was 0.10‰ for $\delta^{13}\text{C}$ and 0.07‰ for $\delta^{15}\text{N}$ across all runs.

Isotopic characterization of salmon shark habitats

We conducted a literature search and collected published stable isotope ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) values [1-33] for known salmon shark prey identified in the literature [34-38] and from

unpublished data (K. Goldman unpublished, D. Parkyn unpublished) (Table S2), and grouped them based on the ecoregion within which the samples were collected. We combined all prey values into prey groups so that every known prey item was represented either at the species, genus, or family level. This list of prey was as comprehensive and inclusive as possible while only including isotopic values that are representative of known salmon shark prey that might be consumed in each ecoregion. By being more inclusive rather than exclusive, we increased the error associated with regional estimates, resulting in more conservative estimates of regional values. When there were multiple studies for a particular prey group, we aggregated the values from the multiple studies into a single mean \pm SD value for that prey group by resampling 2000 values from each prey $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ distribution in the group and combining them into a cumulative distribution for that prey group. The mean \pm SD of the cumulative distribution was then used for that prey group (Table S3). Prey groups within an ecoregion were then combined using the same approach to estimate a mean $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ value for that ecoregion.

Assumption of salmon sharks being generalists

The use of mean regional values as sources in the mixing model assumes that, over the annual time frame that vertebral annuli integrate dietary information, salmon sharks are generalists in each ecoregion and throughout their ontogeny. Salmon sharks are known to be highly opportunistic predators. One of the common descriptions of their diet across their range is “opportunistic” [34; 36], as they focus on various locally abundant prey, although focus on particular prey species (e.g. salmon, squid) does change seasonally and across their range.

In the central and eastern North Pacific where this study occurs, there are only two quantitative studies of salmon shark diet, and they are of very limited seasonal, ontogenetic, and spatial scope. Kubodera et al. [38] provided a snapshot of the diet of juvenile salmon sharks in

the central North Pacific during April and May of 1999-2000, during which they fed primarily upon cephalopods. Hulbert et al. [37] provided a snapshot of adult diet in Prince William Sound, Alaska during the summer salmon runs of July and August 1999-2000 when sharks aggregate to feed on salmon gathering at the mouths of their natal rivers prior to spawning. Not surprisingly, Hulbert's et al. (2005) study found that salmon sharks primarily fed upon aggregating salmon. However, this focus on salmon in Alaska appears to be seasonal. Although they could not identify prey species, Carlisle et al. [39] found that the trophic ecology of salmon sharks shifts seasonally as prey availability and abundance changes, and that the importance of salmon appears to decrease outside the summer salmon runs. Other than these two studies, in the published literature there are only lists of prey species with no quantitative dietary information. There are no published data on the diet of salmon sharks of any age classes from the California Current or Subtropical Gyre.

Due to the lack of dietary information from across their ontogeny, range, and across all seasons, an assumption that salmon sharks are generalists, for which there is evidence, is the most parsimonious and reasonable assumption. Even in the instances where there are some dietary data available, it does not make sense to attempt to integrate this information into an ontogenetic analysis that spans multiple ecoregions when the data only pertain to part of a season, a particular ecoregion, and to particular age classes, especially when we are analyzing vertebral annuli that integrate dietary information over the course of a year.

If there were quantitative dietary data available across more of their range and ontogeny and from a temporal scale pertinent to the time frame at which vertebral annuli integrate dietary information, this information could be used to provide a more refined isoscape as it would reduce the error around mean regional values, potentially greatly. However, in the absence of such data,

an assumption of sharks being generalists, for which there is evidence, is the most conservative approach.

Assumption that vertebral annuli integrate diet over a year

Goldman & Musick (2006) determined that salmon sharks deposit growth bands annually. Therefore, like other accretionary structures such as otoliths, the tissue deposited within a particular band should only reflect diet and habitat during the period of time integrated by that band, which for salmon sharks is a year. This assumes that the growth bands are metabolically inert and there is no metabolic reworking. There is no published evidence that we are aware of that suggests or supports a hypothesis that the corpus calcarea of elasmobranch vertebrae are not metabolically inert, and there is good evidence that they are indeed inert structures. Using bomb radiocarbon to age and validate periodicity of growth band formation, Campana et al. (2002) reported that in the Porbeagle shark (*Lamna nasus*), the salmon shark's congener, vertebral annuli are also deposited annually and that once deposited the "vertebral growth bands appear to be temporally and metabolically stable". This supports our assertion that a salmon shark's vertebra is a metabolically inert accretionary structure that records dietary information over the course of its life. Hence, the tissue within a growth band represents an integrated estimate of diet and habitat over the course of that year of life.

Although the growth bands are metabolically inert, the rate of tissue deposition within a growth band does change with age. However, this translates into variation in the width of growth bands, with young fast growing sharks depositing greater amounts of tissues and having wider growth bands than slower growing older sharks, which deposit very narrow growth bands (see Figure S1). This change in growth band width is why we were unable to collect tissue for older year classes of sharks, their growth bands were simply too narrow to collect enough tissue for

analysis. However, the rate of deposition within an annulus is not important when we are collecting tissue from an entire annulus, which accretes over the period of a year.

Isotopic characterization of oceanic and neritic nurseries

We used discriminant analysis to assign age 0 sharks to either the North Pacific Transition Zone (NPTZ) or California Current nursery area. To do this, we used age 1 and age 2 sharks from adult vertebrae to represent the NPTZ nursery area and the vertebrae of juvenile salmon sharks collected in the California Current to represent the California Current nursery area. We used mixing model results and the distribution of these two groups of sharks relative to the mean ecoregion values to help inform our use of these groups to characterize the different nursery areas.

As is apparent in figure S4, there is no overlap between the age 1 and 2 sharks we used to characterize the NPTZ nursery and neritic ecoregion values. Similarly, the California Current juveniles do not overlap with oceanic ecoregions (with the exceptions that we mention in the following paragraph). This indicates that these two isotopically distinct groups of sharks represent different nursery habitats, one that is more oceanic and one that is more neritic.

We used age 1 and 2 fish to isotopically characterize the NPTZ nursery habitat because the mixing model results indicated a very high (~90%) contribution from oceanic (SAGTZ & STG) resources (Figure 3, S4), indicating use of an oceanic nursery habitat. We believe this oceanic nursery habitat can reasonably be assumed to be the NPTZ, the only described oceanic nursery for salmon sharks [36; 40]. The first several year classes had some $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values that were not well bounded by any particular region but generally in between SAGTZ and STG (Figure 2). We interpreted these low $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values as reflecting use of the NPTZ nursery area, which we were unable to isotopically characterize as an ecoregion due to lack of prey data.

We used juvenile sharks sampled within the California Current to isotopically characterize the California Current nursery area. The fact that these juvenile sharks are isotopically similar to neritic resources (Figure S4) and are distinct from the other “NPTZ nursery” sharks (age 1 and 2 sharks) suggests that these California Current juveniles do indeed reflect increased use of the California Current nursery area relative to the age 1 and 2 sharks. If juvenile sharks migrated into California from oceanic habitats prior to collection, they should isotopically resemble oceanic resources and have lower $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values relative to the California Current juveniles. This was indeed the case for three of the California Current juveniles. In figure S4, there are several California Current juvenile sharks (open triangles) that overlap the NPTZ juvenile sharks (open diamonds), which we interpreted as indicating that these sharks were recent immigrants from oceanic habitats.

Potential influence of ontogenetic changes in trophic level

Understanding how foraging in regions with differences in baseline $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ and shifts in trophic level interact can be challenging when interpreting stable isotope results. Because the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of consumers increases in a consistent manner through successive trophic levels due to trophic discrimination (fractionation) [41], understanding when shifts in trophic level may occur, whether ontogenetically, seasonally, or geographically, is necessary in order to correctly interpret stable isotope results [42]. Ontogenetic shifts in trophic level are nearly universal in fishes [43-45], though shifts in elasmobranchs are generally smaller than those in teleosts because elasmobranchs exhibit direct development which limits the overall growth and shifts in morphology (i.e. gape width) relative to teleosts [46]. Sharks start out life as large predators relative to teleosts, which may begin life as planktivores and transition to becoming predators as they grow, undergoing a clear ontogenetic increase in trophic level [46].

Shifts in trophic level are likely relatively unimportant for salmon sharks, over the spatial and temporal scale of this study, as they do not appear to undergo a significant ontogenetic shift in trophic level. This suggests that observed shifts in stable isotope values are primarily related to changes in distribution (i.e. differences in baseline stable isotope values between regions) and resulting changes in diet, and not trophic level. Adult salmon sharks primarily consume mid-trophic level prey, as do juvenile salmon sharks [34-37]. For example, in the eastern North Pacific, female salmon sharks may seasonally consume large numbers of salmon [34; 37], though they primarily feed upon pink (*Oncorhynchus gorbuscha*, trophic level (TL) 4.1), chum (*Oncorhynchus keta*, TL 3.9), and sockeye salmon (*Oncorhynchus nerka*, TL 4.4) [47]. Juvenile salmon sharks feed extensively upon gonatid squid (TL 4.4 or 4.1 for large gonatid squid and micronekton squid respectively) at certain times of the year [38; 47]. Some minor dietary items may be at a higher trophic level (e.g. Pacific pomfret, *Brama japonica*, TL 5.0) [47], but overall salmon sharks primarily consume mid-trophic level prey.

In addition, the largest shift in stable isotope values occurs at intermediate ages (i.e. age 6-9, Fig. 2b and c), at which point salmon sharks have already attained a large size (>160 cm PCL) and thus would not be limited by gape width, suggesting that shifts in diet related to increasing size are not driving this difference. Therefore, we believe that any potential ontogenetic change in trophic level is less important than changes in habitat. This assumption is supported by the growing body of literature that demonstrates that changes in baseline isotopic values are primarily what drive changes in the isotopic composition of migratory marine species, not changes in trophic level as has often been suggested [48-50]. Hence, the change associated with movements between isotopically distinct food webs is likely the primary driver influencing a migratory consumer's isotopic composition.

Supplemental tables

ID	Sex	Date	PCL (cm)	Missing	Oldest band	Birth
1	F	08/03/07	192	-	11	y
2	F	07/30/07	184	-	12	y
3	F	07/30/07	193	10	11	y
4	F	07/15/07	188	-	10	y
6	F	07/30/07	190	-	10	y
8	F	07/15/07	194	U	12	n
10	F	08/03/07	189	-	10	y
12	F	07/19/07	192	8	11	y
15	F	07/20/07	200	-	10	y
17	F	07/07/07	187	-	9	y
19	F	07/08/07	179	1, 8	12	y
20	F	07/11/07	193	-	12	y
21	F	07/07/07	184	-	8	y
24	F	07/11/07	185	-	12	y
27	F	08/21/07	185	-	12	y
28	F	07/07/07	175	-	11	y
29	F	07/10/07	199	7	10	y
31	M	06/28/09	168	-	14	y
33	F	07/10/07	185	-	9	y
34	M	06/27/09	150	U, 12	13	n

Table S1: Twenty salmon sharks (18 F, 2 M) sampled from Prince William Sound, Alaska in 2007 and 2009. “PCL” is precaudal length, “Missing” denotes bands with no data due to inadequate amount of sample, and “Oldest band” is oldest annulus that we were able to collect tissue from, which does not necessarily equate to maximum age.

Common name	Species	Family or Order
Arrowtooth flounder	<i>Atheresthes stomias</i>	Pleuronectidae
Atka mackerel	<i>Pleurogrammus monoptygius</i>	Hexagrammidae
Barracudina		Paralepididae
Berry armhook squid	<i>Gonatus berryi</i>	Gonatidae
Boreal clubhook squid	<i>Onychoteuthis borealijaponica</i>	Onychoteuthidae
Boreopacific gonate squid	<i>Gonatopsis borealis</i>	Gonatidae
Capelin	<i>Mallotus villosus</i>	Osmeridae
Chinook salmon	<i>Oncorhynchus tshawytscha</i>	Salmonidae
Chiroteuthid squid	<i>Chiroteuthis calyx</i>	Chiroteuthidae
Chub mackerel	<i>Scomber japonicus</i>	Scombridae
Chum salmon	<i>Oncorhynchus keta</i>	Salmonidae
Codfishes		Gadidae
Coho salmon	<i>Oncorhynchus kisutch</i>	Salmonidae
Cranchiid squid	<i>Belonella borealis</i>	Cranchiidae
Daggerteeth	<i>Anopterus pharao</i>	Anopteridae
Dogfish	<i>Squalus suckleyi</i>	Squalidae
Eulachon	<i>Thaleichthys pacificus</i>	Osmeridae
Fiery armhook squid	<i>Gonatus pyros</i>	Gonatidae
Gonatid squid	<i>Gonatus</i> spp.	Gonatidae
Humboldt squid	<i>Dosidicus gigas</i>	Ommastrephidae
Lancetfish	<i>Alepisaurus ferox</i>	Alepisauridae
Lanternfish		Myctophidae
Lumpfishes		Cyclopteridae
Market squid	<i>Doryteuthis opalescens</i>	Loliginidae
Pacific hake	<i>Merluccius productus</i>	Merlucciidae
Pacific herring	<i>Clupea pallasii</i>	Clupeidae
Pacific pomfret	<i>Brama japonica</i>	Bramidae
Pacific saury	<i>Cololabis saira</i>	Scomberesocidae
Pink salmon	<i>Oncorhynchus gorbuscha</i>	Salmonidae
Rockfish	<i>Sebastes</i> spp.	Sebastidae
Sablefish	<i>Anoplopoma fimbria</i>	Anoplopomidae
Sardine	<i>Sardinops sagax</i>	Clupeidae
Sculpins		Cottidae
Shortarm gonate squid	<i>Gonatus middendorfi</i>	Gonatidae
Shrimp		Decapoda
Smallfin gonate squid	<i>Berryteuthis anonychus</i>	Gonatidae
Sockeye salmon	<i>Oncorhynchus nerka</i>	Salmonidae
Squid		Teuthoidea
Steelhead	<i>Oncorhynchus mykiss</i>	Salmonidae
Tanner crab	<i>Chionecetes</i> sp.	Oregoniidae
Tomcod	<i>Microgadus proximus</i>	Gadidae
Tuberculate pelagic octopus	<i>Ocythoe tuberculata</i>	Ocythoidea
Walleye pollock	<i>Theragra chalcogramma</i>	Gadidae

Table S2: List of known salmon shark prey based on Nagasawa (1998), Compagno (2001), Hulbert et al. (2005), Kubodera et al. (2007), Goldman and Musick (2008), and K. J. Goldman (Humboldt squid, unpublished) and D. Parkyn (Pacific hake, unpublished, Program of Fisheries and Aquatic Sciences, University of Florida, Gainesville, FL, USA).

Age	Carbon ($\delta^{13}\text{C}$)			Nitrogen ($\delta^{15}\text{N}$)			N
	Mean	SD	Range	Mean	SD	Range	
U	-15.02	0.52	2.07	15.06	0.62	2.66	18
0	-15.88	0.65	2.52	13.88	1.17	4.72	20
1	-16.47	0.38	1.46	12.94	1.21	4.35	19
2	-16.50	0.66	2.97	12.73	1.13	3.86	20
3	-16.39	0.42	1.59	12.66	1.10	3.66	20
4	-16.28	0.52	1.71	12.75	1.32	4.55	20
5	-16.45	0.69	2.99	12.95	1.35	4.76	20
6	-16.23	0.56	2.63	13.09	0.99	4.76	20
7	-15.86	0.61	2.29	13.63	1.03	4.34	19
8	-15.65	0.57	2.71	13.91	0.63	2.53	18
9	-15.54	0.41	1.82	14.39	0.66	2.31	19
10	-15.64	0.26	0.88	14.33	0.59	2.44	16
11	-15.54	0.42	1.42	14.76	0.63	2.07	12
12	-15.72	0.45	1.47	14.43	0.38	1.19	7
13	-15.60	0.48	0.95	14.66	0.04	0.09	2
14	-15.60	na	na	15.10	na	na	1

Table S3: Summary statistics for all year classes of vertebral isotope data.

Region	Prey_group	Lowest taxonomic ID	Carbon ($\delta^{13}\text{C}$)		Nitrogen ($\delta^{15}\text{N}$)		Tissue		
			Mean	SD	Mean	SD	type	N	References
AK	Arrowtooth flounder	<i>Atheresthes stomias</i>	-17.8	0.1	14.7	0.2	2	17	1
AK	Atka mackerel	<i>Pleurogrammus monopterygius</i>	-19.8	0.2	11.6	0.1	1	40	1, 2
AK	Capelin	<i>Mallotus villosus</i>	-19.8	0.5	11.7	0.3	1	154	3, 4, T.C. Kline unpublished
AK	Chinook salmon	<i>Oncorhynchus tshawytscha</i>	-18.5	0.2	14.8	0.2	2	43	1, 5, 6, 7
AK	Chum salmon	<i>Oncorhynchus keta</i>	-20.9	0.3	11.2	0.4	1	55	5, 6, 7, 8
AK	Coho salmon	<i>Oncorhynchus kisutch</i>	-19.6	0.2	13.6	0.2	2	68	1, 5, 6, 7, 8
AK	Eulachon	<i>Thaleichthys pacificus</i>	-17.8	0.2	13.7	0.2	2	82	3, 4, 9, 10, T.C. Kline unpublished
AK	Goniatid squid	Gonatiidae	-22.0	0.9	13.4	0.7	1	2	11
AK	Magister armhook squid	<i>Berryteuthis magister</i>	-19.2	0.2	13.1	0.4	2	230	3, 12, T.C. Kline unpublished
AK	Market squid	<i>Loligo opalescens</i>	-17.3	0.6	13.9	0.3	1	5	9
AK	Myctophidae	<i>Stenobrachius leucopsarus</i>	-19.3	0.4	13.2	0.5	3	60	13
AK	Pacific cod	<i>Gadus macrocephalus</i>	-17.7	0.3	14.6	0.3	1	31	1, 14
AK	Pacific herring	<i>Clupea pallasii</i>	-19.2	0.2	13.0	0.2	2	393	1, 4, 9, 15
AK	Pink salmon	<i>Oncorhynchus gorbuscha</i>	-21.4	0.4	11.4	0.4	1	78	5, 6, 7, 8, 16
AK	Rockfish	<i>Sebastes</i> spp.	-17.9	0.3	14.4	0.6	1	162	3, 11, 17
AK	Sockeye salmon	<i>Oncorhynchus nerka</i>	-21.3	0.3	11.1	0.3	1	95	5, 7
AK	Walleye pollock	<i>Theragra chacogramma</i>	-19.2	0.1	12.6	0.2	2	1293	1, 3, 4, 15, T.C. Kline unpublished
CA	Chinook salmon	<i>Oncorhynchus tshawytscha</i>	-16.8	0.3	14.0	0.2	1	159	18, 19
CA	Chum salmon	<i>Oncorhynchus keta</i>	-19.5	0.3	12.6	0.6	1	2	18
CA	Coho salmon	<i>Oncorhynchus kisutch</i>	-19.3	1.0	12.9	0.6	1	87	18
CA	Goniatid squid	Gonatopsis spp.	-18.8	0.2	13.3	0.3	1	9	D.J. Madigan unpublished
CA	Humboldt squid	<i>Dosidicus gigas</i>	-17.2	0.2	15.0	0.3	1	41	21, 23, D.J. Madigan unpublished
CA	Market squid	<i>Loligo opalescens</i>	-17.0	0.4	13.5	0.4	2	135	18, 19, 20, 21, D.J. Madigan unpublished
CA	Myctophidae	Myctophidae ^a	-18.8	0.2	14.1	0.3	1	16	22, 23, D.J. Madigan unpublished
CA	Onychoteuthid squid	<i>Onychoteuthis</i> sp.	-18.5	0.1	14.4	0.3	1	6	21
CA	Pacific hake	<i>Merluccius productus</i>	-17.2	0.2	13.7	0.2	1	6	18
CA	Pacific herring	<i>Clupea pallasii</i>	-16.7	0.4	13.1	0.4	1	68	18
CA	Pacific mackerel	<i>Scomber japonicus</i>	-18.1	0.4	14.3	0.3	1	23	18, 21, D.J. Madigan unpublished
CA	Pacific saury	<i>Cololabis saira</i>	-19.4	0.3	12.7	0.4	1	71	11, 18, 21, D.J. Madigan unpublished
CA	Rockfish	<i>Sebastes</i> spp. ^b	-18.5	0.2	12.6	0.2	2	55	18, 19, 20, 24, D.J. Madigan unpublished
CA	Sablefish	<i>Anoplopoma fimbria</i>	-19.0	0.2	14.2	0.4	1	23	18, 19, 22
CA	Sardine	<i>Sardinops sagax</i>	-17.7	0.2	12.9	0.2	2	95	18, 19, 20, 21, 25, 26, D.J. Madigan unpublished
CA	Spiny dogfish	<i>Squalus acanthias</i>	-17.5	0.4	12.6	0.4	1	15	18
CA	Steelhead	<i>Oncorhynchus mykiss</i>	-16.6	1.0	13.4	0.8	1	4	18
CA	Tomcod	<i>Microgadus proximus</i>	-18.2	0.3	12.4	0.1	1	2	18
SAGTZ	Chinook salmon	<i>Oncorhynchus tshawytscha</i>	-19.0	0.6	13.6	0.4	1	13	27, 28
SAGTZ	Chum salmon	<i>Oncorhynchus keta</i>	-21.5	0.6	11.3	0.9	1	55	27, 28
SAGTZ	Coho salmon	<i>Oncorhynchus kisutch</i>	-20.1	0.6	11.7	0.4	1	49	27, 28
SAGTZ	Goniatid squid	Gonatiidae	-21.7	0.5	14.0	1.3	1	2	D.J. Madigan unpublished
SAGTZ	Myctophidae	Myctophidae ^c	-21.4	0.2	11.0	0.5	2	779	13, 29, D.J. Madigan unpublished
SAGTZ	Neon flying squid	<i>Ommastrephes bartrami</i>	-18.4	0.7	11.7	1.3	1	44	29
SAGTZ	Pacific saury	<i>Cololabis saira</i>	-19.7	0.6	10.8	1.0	2	16	29, D.J. Madigan unpublished
SAGTZ	Pink salmon	<i>Oncorhynchus gorbuscha</i>	-20.8	0.7	10.6	0.5	1	57	27, 28
SAGTZ	Pacific pomfret	<i>Brama japonica</i>	-19.2	0.8	10.9	2.1	3	10	29
SAGTZ	Sockeye salmon	<i>Oncorhynchus nerka</i>	-20.3	0.5	11.3	0.4	1	45	27, 28
SAGTZ	Squid (miscellaneous)	Gonatiidae, Octopoteuthidae, Histioteuthidae, Cranchiidae ^d	-18.5	0.4	11.6	2.1	1	5	29
SAGTZ	Steelhead	<i>Oncorhynchus mykiss</i>	-19.0	1.0	12.5	1.0	1	35	27
STG	Miscellaneous forage fish	Miscellaneous (primarily <i>Cololabis saira</i>)	-20.5	0.4	8.4	3.4	1	12	D.J. Madigan unpublished
STG	Myctophidae	Myctophidae	-21.2	0.4	9.5	1.8	1	2	D.J. Madigan unpublished
STG	Neon flying squid	<i>Ommastrephes bartrami</i>	-20.5	0.6	12.4	1.9	1	165, 21 ^f	30, 31
STG	Ommastrephid squid	Ommastrephidae ^e	-18.8	0.1	6.2	0.6	1	2	32, 33
STG	Purpleback flying squid	<i>Sthenoteuthis oualaniensis</i>	-18.3	0.6	10.5	2.6	1	167, 25 ^g	30, 31, D.J. McCauley unpublished ^h

Table S4: Isotopic composition of known prey taxa from the literature used to estimate mean ecoregion $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values used in MixSIR to assess proportional contribution of different regions to salmon shark vertebral tissue. Only studies that accounted for lipid through chemical extraction or mathematical correction were included. Tissues are 1: muscle, 2: muscle and whole organism, 3: whole organism. A) *Stenobrachius leucopsarus*, *Benthoosema panamense*; b) *Sebastes jordani*, *S. rufus*, *S. mystinus*, *S. pinniger*, *S. crameri*, *S. entomelas*; C) *Stenobrachius leucopsarus*, *Electronia* sp., *Symbolophorus* sp., *Lampanyctus* sp., *Diaphus* sp., *Notoscopelus* sp., *Protomyctophum thompsoni*, *Tarletonbeania crenularis*, *Diaphus theta*; D) *Berryteuthis anonychus*, *Octopoteuthis deletron*, *Histioteuthis dofleini*, *Taonius pavo*; E) *Ommastrephes* sp., *Hyaloteuthis* sp.; F) Nitrogen based on 167 samples, carbon 25; G) Nitrogen based on 165 samples, carbon 21; h) Douglas McCauley, Department of Ecology, Evolution and Marine Biology, University of California Santa Barbara, Santa Barbara, CA, USA.

	Carbon ($\delta^{13}\text{C}$)		Nitrogen ($\delta^{15}\text{N}$)	
	Mean	SD	Mean	SD
Alaska	-19.34	1.44	13.05	1.28
California	-18.04	1.04	13.43	0.84
Subarctic Gyre	-19.96	1.29	11.77	1.57
Subtropical Gyre	-19.84	1.19	9.38	2.44

Table S5: Estimates of the mean $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of ecoregions used in MixSIR, values are based on prey values in Table S4.

Supplemental figures

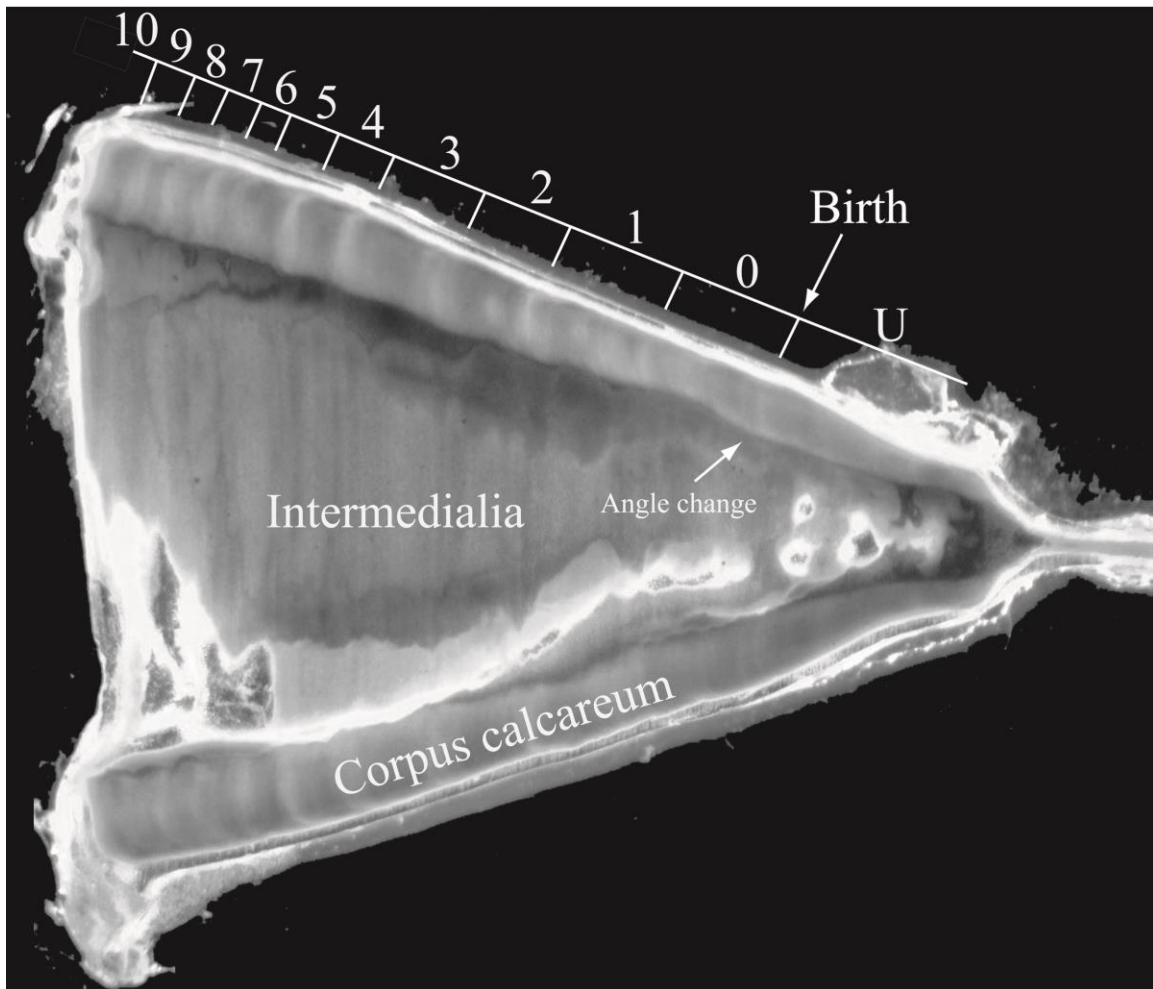


Figure S1: Sagittal section of 10 year old salmon shark vertebrae showing annuli, modified from Goldman and Musick (2006). The birth mark is identified by the angle change. U represents in-utero growth of salmon shark.

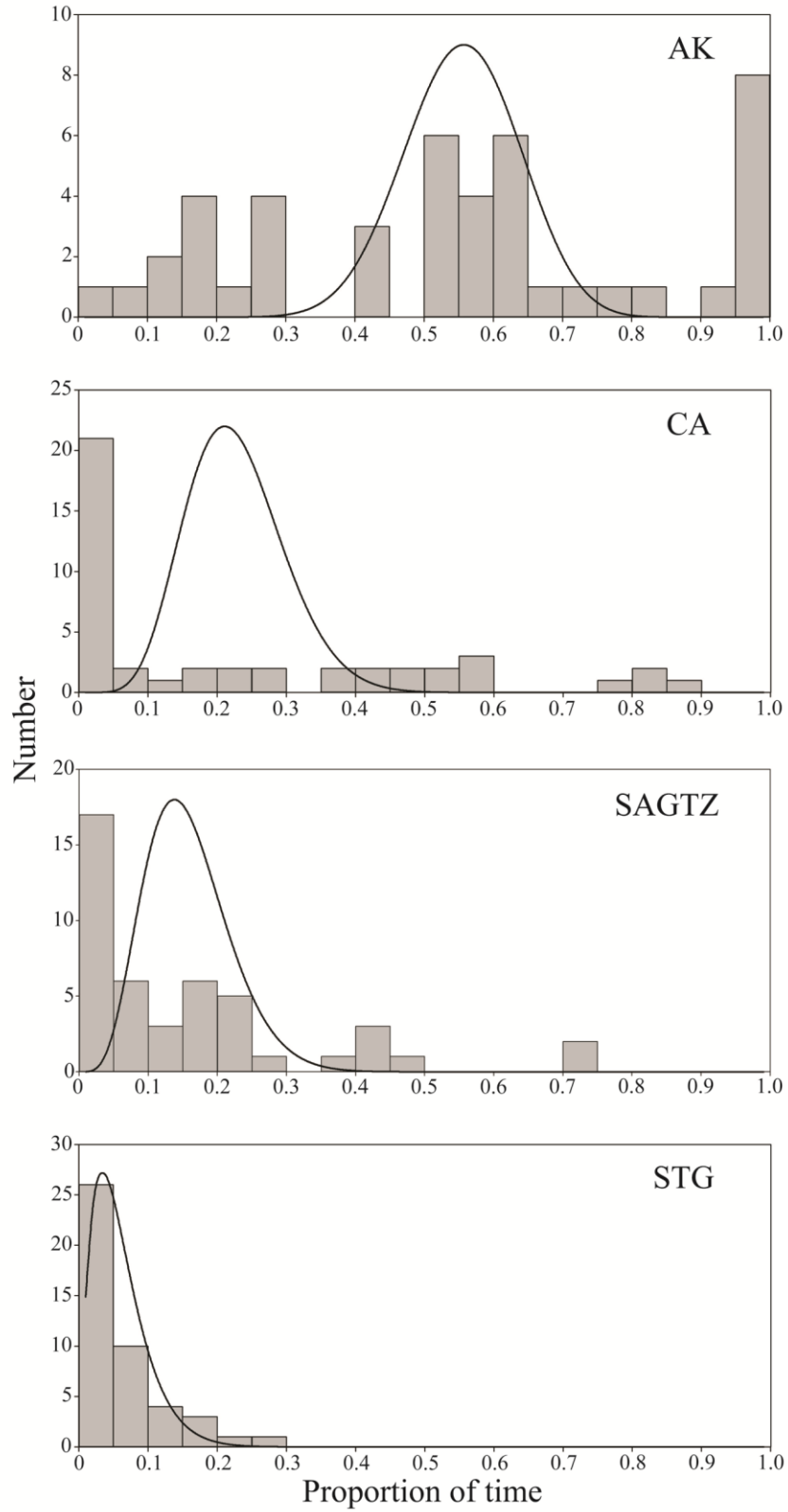


Figure S2: Annual proportion of time spent in the different ecoregions from 44 yearlong tracks of satellite tagged salmon sharks (ages 8+). Lines indicate the specified prior distribution for the contribution of each ecoregion, based on satellite tag data, used in mixing models.

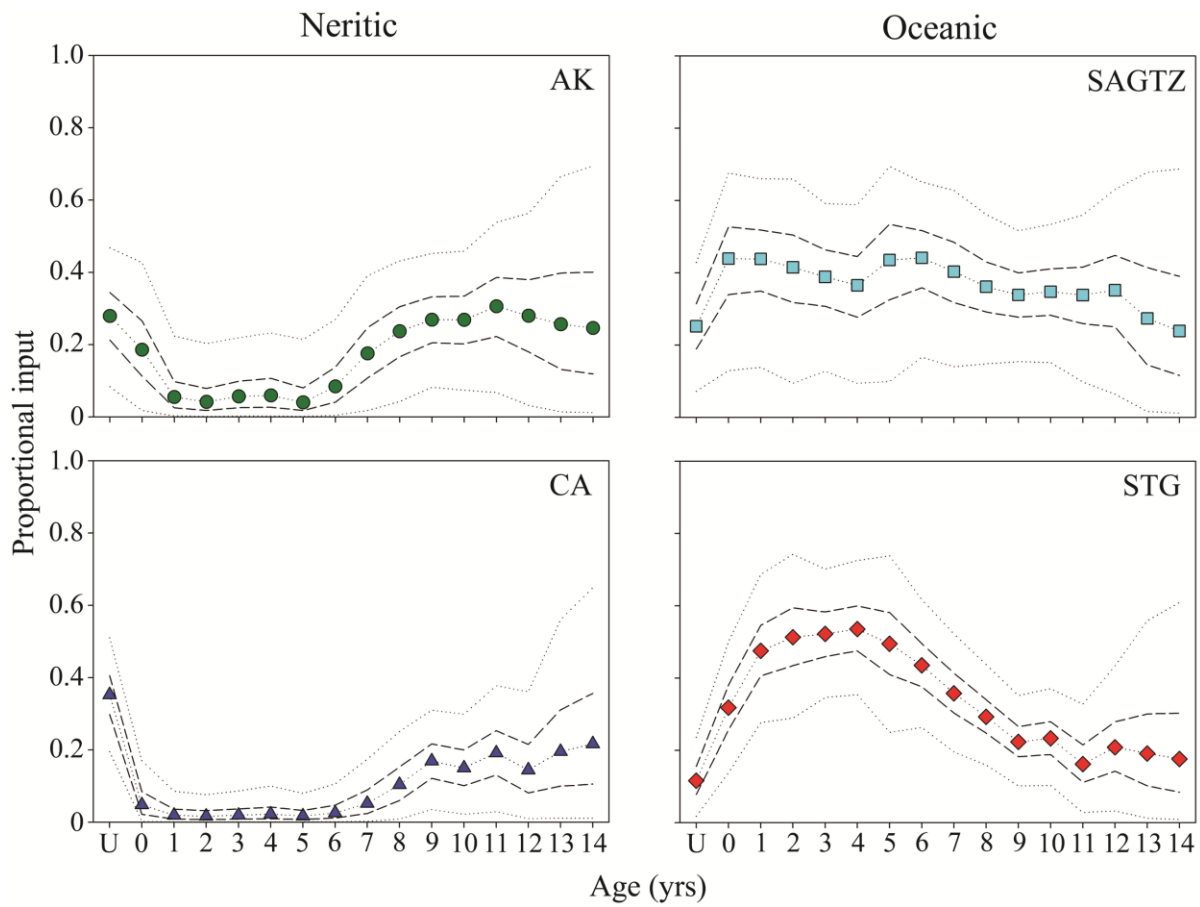


Figure S3: Ontogenetic time series of proportional contribution of the different ecoregions to salmon shark vertebral tissue based on MixSIR using uninformative priors. Median (symbols), interquartile range (dashed lines), and 95% credible intervals (dotted lines) are shown. Note that the left column shows ecoregions with neritic habitats (AK, CA), while the right column shows oceanic ecoregions (SAGTZ, STG). Age U reflects in-utero salmon sharks.

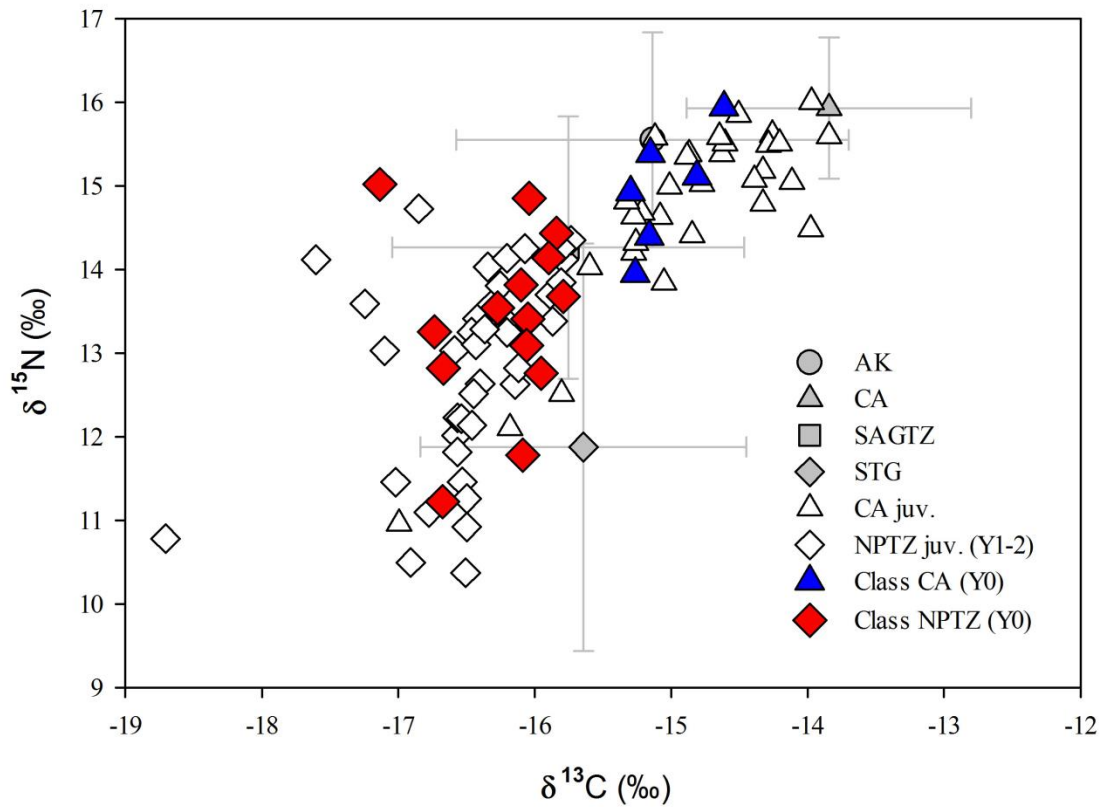


Figure S4: Discriminant analysis results showing classification of year class 0 sharks into either California Current nursery (blue triangle) or NPTZ (red diamond) nursery. Training data (white symbols) used to classify year 0 data into the two groups are shown. CA juveniles are juvenile salmon sharks that were collected in the California Current and represent juveniles that used the California Current as a nursery, while NPTZ juveniles are age 1 and 2 from the vertebral data that represent the NPTZ nursery. Mean (\pm SD) ecoregion estimates (gray symbols) are adjusted to account for trophic discrimination factors.

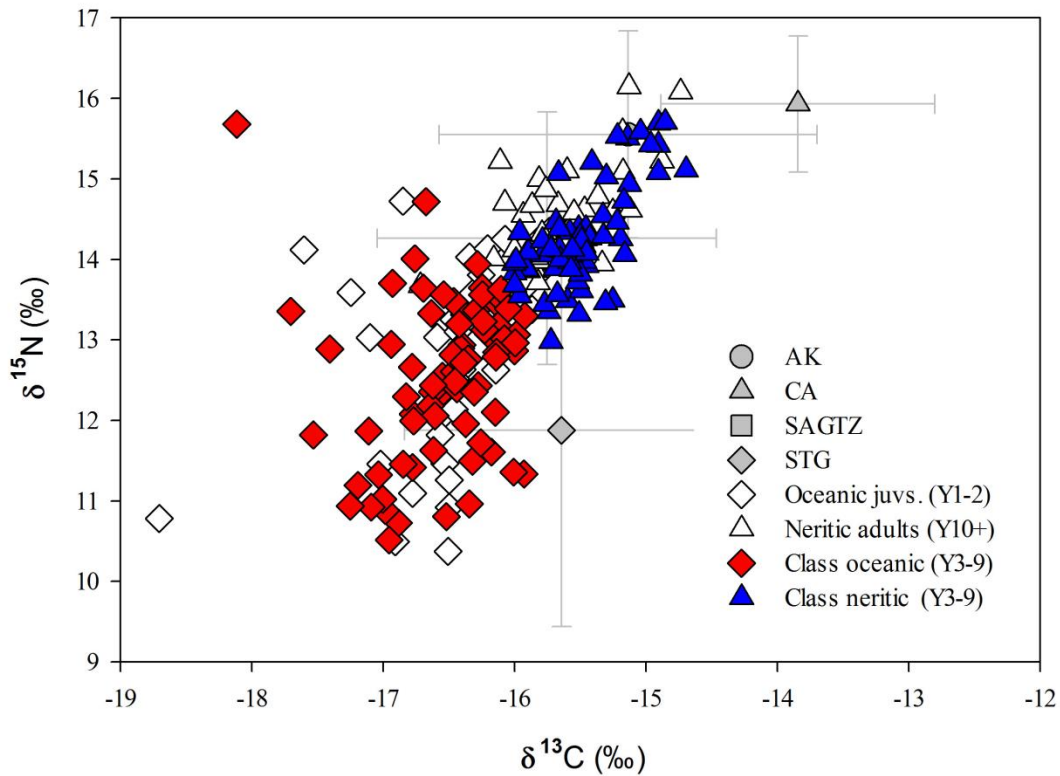


Figure S5: Discriminant analysis results showing classification of age 3 to age 9 sharks into either juvenile habitat (high contribution from oceanic habitats) or adult habitat (increased contribution from neritic habitats). Training data (white symbols) used to classify year 3 – 9 data into the two groups (oceanic, neritic) are shown. Oceanic juveniles are year 1 and 2 sharks that had highest use of juvenile habitats, and neritic adults were large adult sharks (year 10+) that had highest use of neritic habitats. Mean (\pm SD) ecoregion estimates (gray symbols) are adjusted to account for trophic discrimination factors.

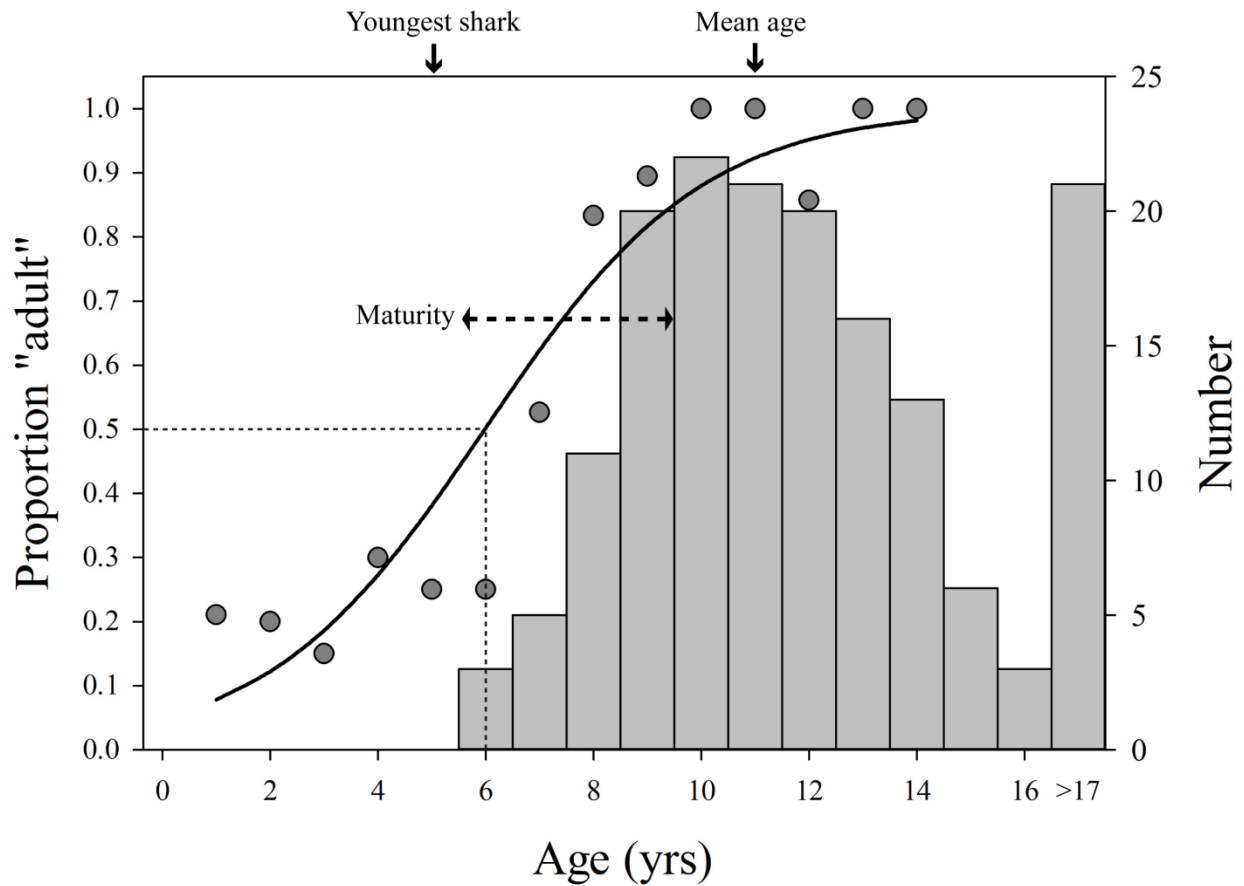


Figure S6: Logistic regression (solid line) of the proportion of sharks (circles) classified as “juvenile” or “adult” using discriminant analysis. The histogram shows estimated ages of salmon sharks ($n = 162$) tagged between 2002 and 2012 by the TOPP program (mean 12.7 yrs \pm 4.1 SD). Minimum (5) and mean (11) age, and age of maturity of sharks ($n = 146$) caught in Alaskan waters and aged by Goldman and Musick (2006) are also shown.

Literature cited

1. Kurle, C. M., Sinclair, E. H., Edwards, A. E. & Gudmundson, C. J. 2011 Temporal and spatial variation in the $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values of fish and squid from Alaskan waters. *Mar Biol* **158**, 2389-2404.
2. Rand, K. M. 2007 Longitudinal growth differences in Atka mackerel (*Pleurogrammus monopterygius*) using a bioenergetic model to identify underlying mechanisms. M.Sc., University of Washington, pp. 58.
3. Kline, T. C. 2007 Rockfish trophic relationships in Prince William Sound, Alaska, based on natural abundance of stable isotopes. In *Biology, assessment, and management of North Pacific rockfishes* (ed. J. Heifetz, DiCosimo, J., Gharrett, A. J., Love, M. S., O'Connell, V. M., Stanely, R. D.), pp. 21-38: Alaska Sea Grant, University of Alaska Fairbanks.
4. Witteveen, B. H., Worthy, G. A. J., Foy, R. J. & Wynne, K. M. 2012 Modeling the diet of humpback whales: An approach using stable carbon and nitrogen isotopes in a Bayesian mixing model. *Mar Mamm Sci* **28**, E233-E250.
5. Johnson, S. P. & Schindler, D. E. 2009 Trophic ecology of Pacific salmon (*Oncorhynchus* spp.) in the ocean: a synthesis of stable isotope research. *Ecol Res* **24**, 855-863.
6. Piorkowski, R. J. 1995 Ecological effects of spawning salmon on several southcentral Alaskan streams. Ph.D., University of Alaska, Fairbanks.
7. Satterfield, F. R. & Finney, B. P. 2002 Stable isotope analysis of Pacific salmon: insight into trophic status and oceanographic conditions. *Prog Oceanogr* **53**, 231 - 246.
8. Ben-David, M. 1996 Seasonal diets of mink and martens: effects of spatial and temporal changes in resource abundance. Ph.D., University of Alaska, Fairbanks, pp. 207.
9. Hobson, K. A., Piatt, J. F. & Pitocchelli, J. 1994 Using stable isotopes to determine seabird trophic relationships. *J Anim Ecol* **63**.
10. Kurle, C. M. 2002 Stable-isotope ratios of blood components from captive northern fur seals (*Callorhinus ursinus*) and their diet: applications for studying the foraging ecology of wild otariids. *Can J Zool* **80**, 902-909.
11. Davies, W. E., Hipfner, J. M., Hobson, K. A. & Ydenberg, R. C. 2009 Seabird seasonal trophodynamics: isotopic patterns in a community of Pacific alcids. *Mar Ecol Prog Ser* **382**, 211-219.
12. Hunsicker, M. E., E., E. T., Aydin, K. Y. & Ishida, B. 2010 Predatory role of the commander squid *Beryteuthis magister* in the eastern Bering Sea: insights from stable isotopes and food habits. *Mar Ecol Prog Ser* **415**, 91-108.
13. Kline, T. C. 2010 Stable carbon and nitrogen isotope variation in the northern lampfish and *Neocalanus*, marine survival rates of pink salmon, and meso-scale eddies in the Gulf of Alaska. *Prog Oceanogr* **87**, 49-60.
14. Ricca, M. A., Miles, A. K., Anthony, R. G., Deng, X. & Hung, S. S. O. 2007 Effect of lipid extraction on analyses of stable carbon and stable nitrogen isotopes in coastal organisms of the Aleutian archipelago. *Can J Zool* **85**, 40-48.
15. Kline, T. C. 1999 Temporal and spatial variability of $^{13}\text{C}/^{12}\text{C}$ and $^{15}\text{N}/^{14}\text{N}$ in pelagic biota of Prince William Sound, Alaska. *Can J Fish Aquat Sci* **56**, 94-117.
16. Chaloner, D. T., Martin, K. M., Wipfli, M. S., Ostrom, P. H. & Lamberti, G. A. 2002 Marine carbon and nitrogen in southeastern Alaska stream food webs: evidence from artificial and natural streams. *Can J Fish Aquat Sci* **59**, 1257-1265.

17. Sorensen, M. C., Hipfner, J. M., Kyser, T. K. & Norris, D. R. 2009 Carry-over effects in a Pacific seabird: stable isotope evidence that pre-breeding diet quality influences reproductive success. *J Anim Ecol* **787**, 460-467.
18. Miller, T. W. 2006 Trophic dynamics of marine nekton and zooplankton in the northern California Current pelagic ecosystem. Ph. D., Oregon State University, pp. 212.
19. Sydeman, W. J., Hobson, K. A., Pyle, P. & McLaren, E. B. 1997 Trophic relationships among seabirds in central California: combined stable isotope and conventional dietary approach. *The Condor* **99**, 327 - 336.
20. Becker, B. H., Peery, M. Z. & Beissinger, S. R. 2007 Ocean climate and prey availability affect the trophic level and reproductive success of the marbled murrelet, an endangered seabird. *Mar Ecol Prog Ser* **329**, 267-279.
21. Madigan, D. J., Carlisle, A. B., Dewar, H., Snodgrass, O. E., Litvin, S. Y., Micheli, F. & Block, B. A. 2012 Stable isotope analysis challenges wasp-waist food web assumptions in an upwelling pelagic ecosystem. *Scientific Reports* **2**, doi:10.1038/srep00654.
22. Boyle, M. D., Ebert, D. A. & Cailliet, G. M. 2012 Stable-isotope analysis of a deep-sea benthic-fish assemblage: evidence of an enriched benthic food web. *J Fish Biol* **80**, 1485-1507.
23. Ruiz-Cooley, R. I., Markaida, U., Gendron, D. & Aguiniga, S. 2006 Stable isotopes in jumbo squid (*Dosidicus gigas*) beaks to estimate its trophic position: comparison between stomach contents and stable isotopes. *J Mar Biol Assoc U K* **86**, 437 - 445.
24. Jarman, W. M., Hobson, K. A., Sydeman, W. J., Bacon, C. E. & McLaren, E. B. 1996 Influence of trophic position and feeding location on contaminant levels in the Gulf of the Farallones food web revealed by stable isotope analysis. *Environ Sci Technol* **30**, 654 - 660.
25. Gendron, D., Aguñiga, S. & Carriquiry, J. D. 2001 $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ in skin biopsy samples: a note on their applicability for examining the relative trophic level in three rorqual species. *J Cetacean Res Manag* **3**, 1-4.
26. Toperoff, A. K. 1997 Diet of harbor porpoise (*P. phocoena*) using stomach contents and stable isotope analyses. MS, San Jose State University, pp. 103.
27. Kaeriyama, M., Nakamura, M., Edpalina, R., Bower, J. R., Yamaguchi, H., Walker, R. V. & Myers, K. W. 2004 Change in feeding ecology and trophic dynamics of Pacific salmon (*Oncorhynchus* spp.) in the central Gulf of Alaska in relation to climate events. *Fish Oceanogr* **13**, 197-207.
28. Welch, D. W. & Parsons, T. R. 1993 ^{13}C and ^{15}N values as indicators of trophic position and competitive overlap for Pacific salmon (*Oncorhynchus* spp.). *Fish Oceanogr* **2**, 11-23.
29. Gould, P., Ostrom, P. & Walker, W. 1997 Trophic relationship of albatrosses associated with squid and large-mesh drift-net fisheries in the North Pacific Ocean. *Can J Zool* **75**, 549 - 562.
30. Parry, M. P. 2003 The trophic ecology of two ommastrephid squid species, *Ommastrephes bartamii* and *Sthenoteuthis oualaniensis*, in the North Pacific sub-tropical gyre. Ph.D., University of Hawaii, pp. 287.
31. Parry, M. P. 2008 Trophic variation with length in two ommastrephid squids, *Ommastrephes bartramii* and *Sthenoteuthis oualaniensis*. *Mar Biol* **153**, 249-256.
32. Graham, B. S., Grubbs, D., Holland, K. & Popp, B. N. 2007 A rapid ontogenetic shift in the diet of juvenile yellowfin tuna from Hawaii. *Mar Biol* **150**, 647-658.

33. Graham, B. S. 2007 Trophic dynamics and movements of tuna in tropical Pacific Ocean inferred from stable isotope analyses. Ph. D. thesis, University of Hawaii, pp. 237.
34. Nagasawa, K. 1998 Predation by salmon sharks (*Lamna ditropis*) on Pacific salmon (*Onchorhynchus* spp.) in the North Pacific Ocean. *North Pac Anadromous Fish Comm Bull* **1**, 419-433.
35. Compagno, L. J. V. 2001 *Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Volume 2. Bullhead, mackerel and carpet sharks (Heterodontiformes, Lamniformes and Orectolobiformes)*. Rome: FAO Species Catalogue for Fishery Purposes No. 1 Vol. 2.
36. Goldman, K. J. & Musick, J. A. 2008 The biology and ecology of the salmon shark, *Lamna ditropis*. In *Sharks of the open ocean: biology, fisheries and conservation* (ed. M. D. Camhi, E. K. Pikitch & E. A. Babcock), pp. 95-104. Blackwell Publishing, Oxford, UK.
37. Hulbert, L. B., Aires-Da-Silva, A. M., Gallucci, V. F. & Rice, J. S. 2005 Seasonal foraging movements and migratory patterns of female *Lamna ditropis* tagged in Prince William Sound, Alaska. *J Fish Biol* **67**, 490-509.
38. Kubodera, T., Watanabe, H. & Ichii, T. 2007 Feeding habits of the blue shark, *Prionace glauca*, and salmon shark, *Lamna ditropis*, in the transition region of the Western North Pacific. *Rev Fish Biol Fish* **17**, 111-124.
39. Carlisle, A. B., Perle, C. R., Goldman, K. J. & Block, B. A. 2011 Seasonal changes in depth distribution of salmon sharks (*Lamna ditropis*) in Alaskan waters: implications for foraging ecology. *Can J Fish Aquat Sci* **68**, 1905-1921.
40. Nakano, H. & Nagasawa, K. 1996 Distribution of pelagic elasmobranchs caught by salmon research gillnets in the North Pacific. *Fisheries Science* **62**, 860-865.
41. Peterson, B. J. & Fry, B. 1987 Stable isotopes in ecosystem studies. *Annu Rev Ecol Syst* **18**, 293-320.
42. Post, D. M. 2002 Using stable isotopes to estimate trophic position: Models, methods, and assumptions. *Ecology* **83**, 703-718.
43. Werner, E. E. & Gilliam, J. F. 1984 The ontogenetic niche and species interactions in size-structured populations. *Annu Rev Ecol Syst* **15**, 393-425.
44. Mittelbach, G. G. & Persson, L. 1998 The ontogeny of piscivory and its ecological consequences. *Can J Fish Aquat Sci* **55**, 1454-1465.
45. Wetherbee, B. M. & Cortes, E. 2004 Food consumption and feeding habits. In *Biology of sharks and their relatives* (ed. J. C. Carrier, J. A. Musick & M. R. Heithaus), pp. 225-246. CRC Press, Boca Raton, FL.
46. Grubbs, R. D. 2010 Ontogenetic shifts in movements and habitat use. In *Sharks and their relatives II: biodiversity, physiology, and conservation* (ed. J. C. Carrier, J. A. Musick & M. R. Heithaus), pp. 319-350. CRC Press, Boca Raton, FL.
47. Aydin, K. Y., McFarlane, G. A., King, J. R. & Megrey, B. A. 2003 The BASS/MODEL report on trophic models of the subarctic basin ecosystems. *North Pacific Marine Science Organization (PICES) Report* **25**, 1-93.
48. Lorrain, A., Graham, B. S., Popp, B. N., Allain, V., Olson, R. J., Hunt, B. P. V., Potier, M., Fry, B., Galván-Magaña, F., Menkes, C. E. R., Kaehler, S. & Ménard, F. 2014 Nitrogen isotopic baselines and implications for estimating foraging habitat and trophic position of yellowfin tuna in the Indian and Pacific Oceans. *Deep Sea Research Part II: Topical Studies in Oceanography*.

49. Popp, B. N., Graham, B. S., Olson, R. J., Hannides, C. C., Lott, M. J., Lopez-Ibarra, G. A., Galvan-Magana, F. & Fry, B. 2007 Insight into the trophic ecology of yellowfin tuna, *Thunnus albacares*, from compound-specific nitrogen isotope analysis of proteinaceous amino acids. In *Stable isotopes as indicators of ecological change* (ed. T. E. Dawson & R. T. W. Siegwolf), pp. 173 - 190. Academic Press, London.
50. Graham, B. S., Koch, P. L., Newsome, S. D., McMahon, K. W. & Aurioles, D. 2010 Using isoscapes to trace the movements and foraging behavior of top predators in oceanic ecosystems. In *Isoscapes: understanding movement, pattern, and process on earth through isotope mapping* (ed. J. B. West, G. J. Bowen, T. E. Dawson & K. P. Tu), pp. 299-318. Springer, New York.